



## The Prevalance, Etiology and Antimicrobial Susceptibility of the Microorganisms in Subclinical Mastitis in Goats\*

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**Abstract:** This study was performed to determine the prevalence, causative microorganisms and antimicrobial susceptibility of microorganisms in goat subclinical mastitis around Hatay. A total of 1010 mammary halves of 505 goats were examined by California Mastitis Test (CMT). The somatic cell counts (SCC) were determined by microscopic method. Isolation and identification of microorganisms were carried out by conventional microbiologic methods. *Staphylococci* were further differentiated by API-STAPH system. Antimicrobial susceptibility was determined by disc diffusion method. The prevalence of the subclinical mastitis was found 8.71 %. The most prevalent microorganism was *staphylococci* (71.5%). Microorganisms except *Staphylococci* were *Streptococci* (8%), *Bacillus spp.* (5.7%), *Escherichia coli* (4.5%), *Corynebacterium spp.* (3.4%), *Pseudomonas spp.* (2.3%) and *Acinetobacteri spp.* (2.3%). In addition, mix infection was defined in 2.3% of samples. Highly resistance was found against penicillin, erythromycin, oxytetracycline, gentamicin, amoxicillin. Slightly resistance was found against enrofloxacin, amoxicillin-clavulonic acid, kanamycin plus cephalexin and there was no resistance against cefalotin. It was concluded that, prevalence of subclinical mastitis should be cared; also *staphylococci* especially *Coagulase negative staphylococci* are the most commonly isolated bacteria in subclinical mastitis around Hatay. In goat mastitis diagnosis strong positive CMT results should be cared. CMT and SCC results should be supported with microbiologic tests.

**Keywords:** Diagnosis, Goat, Prevalence, Subclinic mastitis.

## Keçilerde Subklinik Mastitislerde Prevalans, Etiyoloji ve Mikroorganizmaların Antibiyotik Duyarlılıkları

**Öz:** Çalışma, Hatay çevresinde keçilerde subklinik mastitis prevalansı, neden olan mikroorganizmalar ve mikroorganizmaların antibiyotik duyarlılıklarını belirlemek amacıyla düzenlendi. Beşüzbeş keçiye ait 1010 meme lobu CMT ile muayene edildi. Somatik hücre sayısı (SHS) direkt mikroskopik yöntemle belirlendi. Mikroorganizmaların izolasyon ve identifikasyonu rutin mikrobiyolojik yöntemlerle gerçekleştirildi. Stafilokokların alt türlerinin belirlenmesi amacıyla API-STAPH system kullanıldı. Antibiyotik duyarlılık testlerinde disk difüzyon yöntemi kullanıldı. Subklinik mastitisin prevalansı %8.71 olarak belirlendi. En fazla izole edilen mikroorganizma %71.5 oranıyla Stafilokoklardı. Çalışmada *Stafilokoklar* dışında *Streptokoklar* (%8), *Bacillus spp.* (%5.7), *E. coli* (%4.5), *Corynebacterium spp.* (%3.4), *Pseudomonas spp.* (%2.3) ve *Acinetobacteri spp.* (%2.3) de izole edildi. Ek olarak örneklerin % 2.3'ünde miks enfeksiyon saptandı. Antibiyotik duyarlılık testlerine göre penisilin, eritromisin, oksitetrasiklin, gentamisin, amoksisilin'e karşı yüksek oranda direç gözlemlendi. Enrofloksasin, amoksisilin klavulonik asit, kanamisin ve sefaleksim'e karşı orta düzeyde direnç saptanırken, sefalotine karşı direnç yoktu. Sonuç olarak Hatay çevresinde keçi sürülerinde subklinik mastitisin prevalansının dikkate değer seviyelerde olduğu, koagülaz negatif stafilokokların en fazla izole edilen mikroorganizmalar olduğu belirlendi. Keçilerde subklinik mastitis tanısında kuvvetli CMT reaksiyonlarının dikkate alınması gerektiği, CMT ve somatik hücre sayılarının mikrobiyolojik yöntemlerle desteklenmeleri gerektiği kanısına varıldı.

**Anahtar Kelimeler:** Keçi, Prevalans, Subklinik mastitis, Tanı.

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## INTRODUCTION

Economically, sub-clinical mastitis is more important than clinical mastitis. It usually precedes the clinical form because of its longer duration also difficult to be detected it has adverse effects on milk quality and production also constitutes a reservoir of microorganisms that lead to infection of other animals within the herd (1-4). The prevalence of intramamillar infection in dairy goats is not only economic but also an hygienic and safety issue with respect to the bacteriological quality of milk in the dairy industry (5).

The prevalence of subclinical mastitis in small ruminants averages 6.5 and 40.2% (6-13). The causative etiologic agents of subclinical mastitis are mainly bacterial origin. Members of the genus *Staphylococci* are the main etiological agents involved in all forms of mastitis in goats. *Coagulase-negative staphylococci* (CNS) are the predominant etiologic agents in goats mastitis, especially in subclinical mastitis (14-17). Although CNS are less pathogenic than *S. aureus*, they can also produce persistent subclinical mastitis and even clinical mastitis (18,19), as well as producing thermostable enterotoxins (20-22). In the previous studies the bacteria isolated except the CNS were *Streptococci* (5,11,12), *Bacillus* spp. (12,13), *E. Coli* (5,11,12), *Corynebacterium* spp. (11), *Klepsiella* spp. (11), *Pseudomonas* spp. (5), *Micrococcus* spp. (13), *Candida* and *Yeast* (11).

Bacterial culture, somatic cell counts (SCC) and California mastitis test (CMT) are widely used to diagnose subclinical mastitis in small ruminants (9,23). SCC is the method based on counting of cells which take part in defense of mammary glands and used as the predictor of mammary health (9,10,23). CMT is the indirect and subjective method of estimating the SCC based on scoring the degree of gel formation of milk with the CMT solution (9,24). The reagent used in CMT consists of the detergent sodium alkylarylsulfonate, sodium hydroxide and a pH indicator. The detergent lysis the somatic cells in milk releasing DNA in the solution of sodium

hydroxide causing formation of a gel (24). Mc Dougall et al. (9) found a positive relationship between SCC and CMT therewithal reported that SCC and CMT are related to bacterial infection in goat milk. However, a positive relationship between SCC and mastitis in dairy goats remains controversial. Otherwise, Karzis et al. (25) stated that CMT and SCC were not alone reliable methods the results should be supported with the bacterial culture. Contreras et al. (23) interprets the bacterial culture as the golden standart method for diagnosing mastitis in goats.

This study was performed to determine the prevalence of the subclinical mastitis, to identify the pathogens and, define the antibiotic susceptibility of the pathogens in goats.

## MATERIALS and METHODS

This survey was carried out on 17 commercial dairy goat flocks on 1010 mammary halves of the 505 lactating clinically healthy dairy Damascus and Hair goats in Hatay. During the study local ethic rules were applied. The goats were milked handly once a day also the kids were suckled after milking. No preventive methods for bacterial transmission, such as antiseptics or gloves, were used during the milking process by the milkers. Teat dipping, neither lactation nor dry-off treatment was conducted in the herds before the study.

CMT solution used in cattle practice was also used in this study. The gradings of the CMT examinations were carried out according to Schalm et al. (26).

Milk samples were aseptically taken from separate udder halves and reached to the laboratory in 1 hour. SCC was determined by microscopic method. Ten µl of milk sample was spread on 5X20 mm area on a microscope slide and air dried. It was fixed by may grünwald solution for 5-7 minutes then stained by Giemsa solution (1 drop giemsa stock/1 ml distilled water) for 25 minutes. Twenty area was examined and the cells were counted. The average of the cell number was enumerated. Microscope

working factor was calculated. The SCC in 1 ml of the sample was found by the formula of Average cell number  $\times$  microscope working factor  $\times$  100.

One hundred microliters of milk sample was spread on blood agar plates (supplemented with 5% defibrinated sheep blood). Aerobic incubation at 37 °C was performed and the plates were examined after 24 h. Identification of the colonies were done according to Gram staining, morphologic and hemolytic status. Tube coagulase test was applied to the *Staphylococcus* spp. colonies. The identification of the *staphylococci* were determined by API-STAPH tests.

The antibiotic susceptibility test was carried out by Kirby Bauer disc diffusion test using the following antibiotic discs; penicillin G (10U, Oxoid), amoxicillin alone (25 µg, Oxoid), gentamycin (10 µg, Oxoid), erythromycin (15 µg, Oxoid), oxytetracycline (30 µg, Oxoid), amoxicillin/clavulonic acid (30 µg, Oxoid), trimethoprim-sulphamethoxazole (25 µg, Oxoid), enrofloxacin (5 µg, Oxoid), cephalotine (30 µg, Oxoid,)), kanamycine plus cefalexin, The susceptibility was calculated according to inhibition zone diameters. The inhibition zone diameters were evaluated according to the interpretive standards of Clinical Laboratory Standards Institute (27).

The samples both giving positive CMT reaction and microorganism isolation occurred were mentioned to be mastitis.

## RESULTS

212 milk samples belonging to 1010 mammary halves gave different grades of positive reactions to CMT (20.99%). Of these, microorganism isolation occurred in 88 samples (41.5%). This ratio gave the prevalence of the mastitis as 8.71% of the total samples. The distribution of the 88 microorganism isolated samples according to CMT scores were; 34 samples CMT score 3, 41 samples CMT score 2, 9 samples were CMT score 1 and 4 samples trace. No microorganism isolation occurred in CMT negative samples. Mean SCC of the samples were  $832 \times 10^3$ ,  $834 \times 10^3$ ,  $1\,036 \times 10^3$ ,  $3\,900 \times 10^3$  and  $7\,932 \times 10^3$  in the CMT negative, trace, CMT score 1, CMT score 2, CMT score 3 respectively. The CMT results and microorganism isolation rates were defined in table 1. The SCC and the microorganism isolation rates according to the CMT scores were summarized in table 2.

**Table 1.** The CMT results and microorganism isolation findings.

**Tablo 1.** CMT sonuçları ve mikroorganizma izolasyon bulguları.

Parameter	(n)	(%)
Samples	1010	100
CMT positive samples	212	20.99
Microbiologic isolation positive in total samples	88	8.71
Microorganism isolation in CMT negative samples	0	0
Microorganism isolation in CMT positive samples	88	41.5

CMT: California mastitis test

**Table 2.** The CMT scores, SCC and the microorganism isolation rates.

**Tablo 2.** CMT skorları, SCC ve mikroorganizma izolasyon oranları.

CMT score	Negative	Trace	1	2	3
SCC/ml	$832 \times 10^3$	$834 \times 10^3$	$1.036 \times 10^3$	$3.900 \times 10^3$	$7.932 \times 10^3$
Microbiologic isolation	0	4	9	41	34
No Microbiologic isolation	All	70	47	5	2

CMT: California mastitis test, SCC: Somatic cell count

Most of the isolates (71.5%) were Streptococci (7%), Bacillus spp. (5.7%), E.coli (4.5%), Staphylococci. Other bacteria isolated were Corynebacterium spp. (3.4%), Pseudomonas spp.

(2.3%), and *Acinetobacter* (2.3%). The mix infection rate was 2.3%. The microorganisms isolated from the milk samples are shown in table 3.

**Table 3.** The microorganisms isolated from the milk samples.

**Tablo 3.** Süt örneklerinden izole edilen mikroorganizmalar.

Microorganisms	% (n)
<i>Staphylococcus spp.</i>	71.5 (63)
<i>Streptococcus spp.</i>	8 (7)
<i>Bacillus</i>	5.7 (5)
<i>Escherichia coli</i>	4.5 (4)
<i>Corynebacterium spp.</i>	3.4 (3)
<i>Pseudomonas</i>	2.3 (2)
<i>Acinetobacter</i>	2.3 (2)
<i>Mix infection</i>	2.3 (2)
Total	100 (88)

The majority of the staphylococcal isolate was *S. intermedius* (23.7%) a CPS. The CNS isolates were *S. capitis* (14.3%), *S. haemolyticus* (9.5%), *S. xylois* (7.9%), *S. simulans* (7.9%), *S. caprae* (7.9%), *S. epidermidis* (6.4%), *S. warneii* (6.4%). Other species were *S. sciuri* (4.8%), *S. hominis* (3.2%), *S. auricularis* (3.2%). *Staphylococcus* species isolated from the milk samples in the study are demonstrated in table 4.

According to the antimicrobial susceptibility test results of the staphylococci; a higher resistance was found against penicilin (77.7%), erythromycin (64.4%), oxytetracycline (53.3%), gentamicin (53.3%), amoxicillin alone (51.1%), the resistance against trimethoprim- sulfamethoxazole was 33.3 %. Slightly resistance was found against enrofloxacin (17.7%), amoxicillin-clavulanic acid (11.1%) and kanamycin plus cephalixin (0.6%) and no resistance was found against cefalotin (0%).

**Table 4.** *Staphylococcus* species identified from the milk samples.

**Tablo 4.** Süt örneklerinden identifiye edilen *Staphylococcus* türleri.

	Species	(n)	%
Coagulase positive	<i>S. aureus</i>	3	4.8
	<i>S. intermedius</i>	15	23.7
Coagulase negative	<i>S. capitis</i>	9	14.3
	<i>S. haemolyticus</i>	6	9.5
	<i>S. xylois</i>	5	7.9
	<i>S. simulans</i>	5	7.9
	<i>S. caprae</i>	5	7.9
	<i>S. epidermidis</i>	4	6.4
	<i>S. warneii</i>	4	6.4
	<i>S. sciuri</i>	3	4.8
	<i>S. hominis</i>	2	3.2
	<i>S. auricularis</i>	2	3.2
	Total	63	100

S.: *Staphylococcus*

## DISCUSSION and CONCLUSION

The prevalence of subclinical mastitis in small ruminants averages 6.5 and 40.2% (7-13). The prevalence of the subclinical mastitis is determined as 8.71% in this study. The range of the prevalence is among the data pointed out in the cited references.

Somatic cells are used as an index of milk quality for cow and goat milk. Milk somatic cell counts of goats are higher than milk somatic cell counts of cows and sheep (28). In goat milk, a sample can be classified as mastitic if it has a SCC of  $1 \times 10^6$  cells per milliliter or greater (29). It is generally agreed in late lactation healthy goats often produce milk with more than one million somatic cells per ml (30-32). The CMT score has been shown to be positively associated with SCC with the probability of bacterial infection (33). Although Perrin et al. (34) reported that, CMT negative (scores 0 and 1) appeared to be more efficient than CMT positive (scores 2 and 3), which probably detects some false reactions which are not related to high SCC in goats, Contreras et al. (33), stated that the CMT Scores 2 and 3 discriminated between infected and uninfected udder glands. Haenlein (35), found that CMT levels in goat milk could determine the infected udder halves. Isolation of bacteria was associated with an

increased SCC, CMT and reduced impedance in both sheep and goats (9). Karzis et al. (25) reported that neither CMT nor SCC was sufficient alone in diagnosis of subclinical mastitis in dairy goats and the results should be approved by microbiological examinations. In the current study majority of the microorganism isolation provided in strong positive (+2 and +3) CMT samples. So, strong positive CMT results found to be predictive for microorganism isolation and mastitis determination, consistent with the findings of Contreras et al. (33). Nevertheless, negative CMT results and slightly positive reactions (trace and 1) should be evaluated carefully and these results should be confirmed by microbiologic examinations for the diagnosis of goat mastitis. Despite the fact that increased number of lactation, lower milk yield, increased number of parturition were reported to increase the SCC in goats without mastitis diagnosis (34,36), in the studies in which the number of parturition, lactation and milk yield was not evaluated and only one sample was taken the CMT results should be supported with the microbiological results as in this research.

Zeng et al. (32), reported that the microscopic method and the Fossomatic machine calibrated with goat milk standards gave comparable results of SCC in goat milk. Based on the CMT results, it was pointed out that the SCC of the samples counted by Fossomatic varied between  $320 \times 10^3$  and  $730 \times 10^3$ ,  $2647 \times 10^3$  and  $6518 \times 10^3$  in the negative trace, 1, 2 and 3, respectively (29). Contreras et al. (33) determined that arithmetical means of SCC per microlitre for each CMT score were,  $312 \times 10^3$  for Score 0 and traces;  $1014 \times 10^3$  and Score 1;  $2912 \times 10^3$  for Score 2; and  $4950 \times 10^3$  for Score 3. In our study, the mean SCC results counted by direct microscopy was obtained  $832 \times 10^3$ ,  $834 \times 10^3$ ,  $1.036 \times 10^3$ ,  $3.900 \times 10^3$  and  $7.932 \times 10^3$  in the negative, trace, 1, 2 and 3 scores of CMT, respectively.

The major types of bacteria involved in sheep and goats are *Staphylococci*, especially various coagulase-negative staphylococci (CNS), that are found on the skin of the udder and its surroundings

(35,37-41). CNS was isolated between the ranges of 34.4% and 95.7%, (11,16,35,42,43). Consistent with the cited references, the major type of bacteria isolated in our study was staphylococci, at a rate of 71.5%, 68.2% of this was CNS. According to the API-STAPH results the CNS types isolated in the current study were; *S. capitis*, *S. haemolyticus*, *S. xylois*, *S. simulans*, *S. caprae*, *S. epidermidis*, *S. warneii*, *S. sciuri*, *S. hominis*, *S. auricularis*. *S. intermedius* coagulase positive staphylococci were the most frequently isolated staphylococci (23.7) in the current study. *S. intermedius* was also isolated among the rates of 9.9 and 26.3% in various studies (22,42).

Da Silva et al. (17), found a resistance in staphylococcus species against penicillin at a rate of 29%, erythromycin and trimethoprim-sulfamethoxazole at a rate of 14%, and the staphylococci was susceptible to oxytetracycline at a rate of 100%. Aydın et al. (11), determined a higher resistance to penicillin, lactams and lactam inhibitors. Moroni et al. (10), reported that both Amoxicillin plus clavulanic acid and Amoxicillin alone showed a good efficacy against *CNS strains*. In the current study a higher resistance to Penicillin, erythromycin, oxytetracycline, gentamicin and amoxicillin alone was determined but a higher susceptibility was observed against amoxycillin plus clavulanic acid. Moroni et al. (10), stated that while cefoperazone showed poor activity against *CNS strains*, cephalonium showed good activity against *CNS strains*. Our results showed that staphylococcus species in the study showed a very high susceptibility against cephalosporins and their combinations such as kanamycin-cephalexim. Aydın et al. (11), showed that staphylococcal species were very susceptible to fluoroquinolone antibiotics as found in this research.

The bacteria isolated except the *staphylococci* were *Streptococci* (5,11,12), *Bacillus spp.* (12,13), *E. Coli* (5,11,12), *Corynebacterium spp.* (11), *Klepsiella spp.* (11), *Pseudomonas spp.* (5), *Micrococcus spp.* (13), *Candida and Yeast* (11). Other bacteria isolated in this study were *Streptococci*, *Bacillus spp.*, *E.coli*,

*Corynebacterium* spp., *Pseudomonas* spp., and *Acinetobacter* spp.

It was concluded that, staphylococci especially the CNS species are the agents most commonly isolated bacteria in subclinical mastitis in the goats herds of Hatay. In the diagnosis of goat mastitis especially the strong positive CMT results should be cared. CMT and SCC results should be supported with microbiologic tests. Also, the differences observed in antimicrobial susceptibility tests, defines the importance of antimicrobial susceptibility testing.

#### REFERENCES

- Hamed Al., Abou-Zeid NA., Kebary KMK., Radwan AA., 1993. Physical and chemical properties of subclinical mastitic sheep and goat milk. *Egyptian Journal of Dairy Science*, 21, 133- 149.
- Urech E., Puhán Z., Schalibaum MS., 1999. Changes in milk protein fraction as affected by subclinical mastitis. *Journal of Dairy Science*, 82, 2402-2411.
- Shearer JK., Harris B., 2003. Mastitis in dairy goats. Animal Science Department Florida Cooperative Extension Service Institute of Food and Agricultural Sciences, 1-6, University of Florida Gainesville, USA.
- Yağcı İP., 2008. Koyunlarda subklinik mastitis: Etiyoloji, epidemiyoloji ve tanı yöntemleri. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi*, 14, 117-122.
- White EC., Hinckley LS., 1999. Prevalance of mastitis pathogens in goat milk. *Small Ruminant Research*, 33, 117-121.
- Aulrich K., Barth K., 2010. Intramammary infections caused by coagulase-negative staphylococci and the effect on somatic cell counts in dairy goats. *Landbauforschung Agriculture and Forestry Research*, 1/2, 59-64.
- Lerondelle C., Poutrel B., 1984. Characteristic of non-clinical mammary infection of the goat. *Annals of Veterinary Research*, 15, 105-112.
- Contreras A., Corrales JC., Sierra D., 1995. Prevalence and aetiology of non-clinical intramammary infection in Murciano-Granadina goats. *Small Ruminant Research*, 17, 71-78.
- McDougall S., Murdough P., Pankey W., Delaney C., Barlow J., Scruton D., 2001. Relationships among somatic cell count, California mastitis test, impedance and bacteriological status of milk in goats and sheep in early lactation. *Small Ruminant Research*, 40, 245-254.
- Moroni P., Pisoni G., Ruffo G., Boettcher PJ., 2005. Risk factors for intramammary infections and relationship with somatic-cell counts in Italian dairy goats. *Prev Vet Med*, 69, 163-173.
- Aydın I., Kav K., Celik HA., 2009. Identification and antimicrobial susceptibility of subclinical mastitis pathogens isolated from hair goats' milks. *Journal of Animal Veterinary Science*, 8, 1086-1090.
- Kostelic A., Cergolj M., Tariba B., Rupic V., Benic M., Gantner V., Stokovic I., 2009. Prevalence and aetiology of subclinical mastitis in goats. *Italian Journal of Animal Science*, 8, 134-136.
- Viridis S., Scarano C., Cossu F., Spanu V., Spanu C., De Santis EPL., 2010. Antibiotic resistance in *Staphylococcus aureus* and coagulase negative staphylococci isolated from goats with subclinical mastitis. *Veterinary Medicine International*, doi: 10.4061/2010/517060.
- Poutrel B., 1984. Udder infection of goats by coagulase-negative staphylococci. *Veterinary Microbiology*, 9, 131-137.
- Bedidi-Madani N., Greenland T., Richard Y., 1998. Exoprotein and slime production by coagulase-negative staphylococci isolated from goats' milk. *Veterinary Microbiology*, 59, 139-145.
- Contreras A., Paape MJ., Miller RH., 1999. Prevalence of subclinical intramammary infection caused by *Staphylococcus epidermidis* in a commercial dairy goat herd. *Small Ruminant Research*, 31, 203-208.
- Da Silva ER., Siqueira AP., Martins JCD., Ferreira WPB., da Silva N., 2004. Identification and in vitro antimicrobial susceptibility of *Staphylococcus* species isolated from goat mastitis in the Northeast of Brazil. *Small Ruminant Research*, 55,

- 45-49.
18. Deinhofer M., Pernthaner A., 1995. Staphylococcus spp. as mastitis-related pathogens in goat milk. *Veterinary Microbiology*, 43,161-166.
  19. Contreras A., Corrales JC., Sanchez A., Sierra D., 1997. Persistence of subclinical intramammary pathogens in goats throughout lactation. *Journal of Dairy Science*, 80, 2815-2819.
  20. Meyrand A., Montet MP., Bavai C., Ray-Gueniot S., Mazuy C., Gaspard CE., Jaubert G., Perrin G., Vernozy-Rozand C., 1999. Risk linked to an enterotoxigenic strain of *Staphylococcus lentus* during the manufacture and ripening of raw goats' milk Camembert-type cheeses. *Revue de Medecine Veterinaire*, 150, 703-708.
  21. Udo EE., Al-Bustan MA., Jacob LE., Chugh TD., 1999. Enterotoxin production by coagulase-negative staphylococci in restaurant workers from Kuwait City may be a potential cause of food poisoning. *Journal of Medical Microbiology*, 48, 819-823.
  22. Contreras A., Luengo C., Sanchez A., Corrales JC., 2003. The role of intramammary pathogens in dairy goats. *Livestock Production Science*, 79, 273-283.
  23. Contreras A., Sierra D., Sanchez A., Corrales JC., Marco JC., Paape MJ., Gonzalo C., 2007. Mastitis in small ruminants. *Small Ruminant Research*, 68, 145-153.
  24. Paape MJ., Poutrel B., Contreras A., Marco JC., Capuco AV., 2001. Milk somatic cells and lactation in small ruminants. *Journal of Dairy Science*, 84, 237-244.
  25. Karzis J., Donkin EF., Petzer IM., 2007. The influence of intramammary antibiotic treatment, presence of bacteria, stage of lactation and parity in dairy goats as measured by the California milk test and somatic cell counts. *Onderstepoort Journal of Veterinary Research*, 74, 161-167.
  26. Schalm OW., Carrol JE., Jain NC., 1971. *Bovine Mastitis* 1<sup>st</sup> ed., 132-153, Lea and Febiger, Philadelphia.
  27. Clinical and Laboratory Standard Institute (CLSI), 2008. Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals, 3<sup>rd</sup> ed., Approved standard M31-A3. Wayne, PA.
  28. Paape MJ., Capuco AV., 1997. Cellular Defense-Mechanisms in the Udder and Lactation of Goats. *Journal of Animal Science*, 75, 556-565.
  29. Poutrel B., Lerondelle C., 1983. Cell content of goat milk: California mastitis test, Coulter counter, and Fossomatic for predicting half infection. *Journal of Dairy Science*, 66, 2575-2579.
  30. Park YW., 1991. Interrelationships between somatic cell counts, electrical conductivity, bacteria counts, percent fat and protein in goat milk. *Small Ruminant Research*, 5, 367-375.
  31. Zeng SS., Escobar EN., 1995. Effect of parity and milk production on somatic cell count, standard plate count and composition of goat milk. *Small Ruminant Research*, 17, 269-274.
  32. Zeng SS., Escobar EN., Hart SP., Hinckley L., Baulthauc M., Robinson GT., Jahnke G., 1999. Comparative study of the effects of testing laboratory, counting method, storage and shipment on somatic cell counts in goat milk. *Small Ruminant Research*, 31, 103-107.
  33. Contreras A., Sierra D., Corrales JC., Sanchez A., Marco J., 1996. Physiological threshold of somatic-cell count and California Mastitis Test for diagnosis of caprine subclinical mastitis. *Small Ruminant Research*, 21, 259-264.
  34. Perrin GG., Mallereau MP., Lenfant D., Baudry C., 1997. Relationships between California mastitis test (CMT) and somatic cell counts in dairy goats. *Small Ruminant Research*, 26, 167-170.
  35. Haenlein GFW., 2002. Relationship of somatic cell counts in goat milk to mastitis and productivity. *Small Ruminant Research*, 45, 163-178.
  36. Wilson DJ., Stewart KN., Sears PM., 1995. Effects of stage of lactation, production, parity and season on somatic cell counts in infected and uninfected dairy goats. *Small Ruminant Research*, 16, 165-169.

37. Bergonier D., De Cremoux R., Rupp R., Lagriffoul G., Berthelot X., 2003. Mastitis of dairy small ruminants. *Veterinary Research*, 34, 689-716.
38. Leitner G., Chaffer M., Zamir S., Mor T., Glickman A., Winkler M., Weisblit L., Saran A., 2000. Udder disease etiology, milk somatic cell counts and NAGase activity throughout lactation in Israeli Assaf sheep. *Small Ruminant Research*, 39,107–112.
39. Leitner G., Chaffer M., Carasso Y., Ezra E., Kababea D., Winkler M., Saran A., 2003. Udder infection and milk somatic cell count, *NAGase* activity and milk composition fat, protein and lactose in Israeli Assaf and Awassi sheep. *Small Ruminant Research*, 49, 157-164.
40. Leitner G., Chaffer M., Shamay A., Shapiro F., Merin U., Ezra E., Saran A., Silanikove N., 2004. Changes in milk composition as affected by subclinical mastitis in sheep. *Journal of Dairy Science*, 87, 46-52.
41. Leitner G., Merin U., Silanikove N., 2004. Changes in milk composition as affected by subclinical mastitis in goats. *Journal of Dairy Science*, 87, 1719-1726.
42. Kalogridou-Vassiliadou D., 1991. Mastitis-related pathogens in goat milk. *Small Ruminant Research*, 4, 203-212.
43. Sung YY., Wu TI., Wang PH., 1999. Evaluation of milk quality of Alpine, Nubian, Saanen and Toggenburg breeds in Taiwan. *Small Ruminant Research*, 33, 17-23.